Purpose: The radial forearm flap is one of the most commonly used methods for intraoral reconstruction in oral carcinoma surgery. One of its disadvantages is the residual functional and unaesthetic defect in the donor site. The objective of this report is to describe preliminary cases of a novel method to cover such donor sites based on the use of autologous skin equivalents (ASEs) and an artificial dermal matrix (Integra, Prim, Barcelona, Spain).

Materials and Methods: The donor sites of 2 patients were treated with the artificial dermal matrix after raising a radial forearm flap. A skin biopsy and a blood sample were taken to construct an ASE. After 3 weeks, the ASE was applied over the dermal template and left to heal. The functional and esthetic results were recorded.

Results: Good functional and esthetic results were achieved, with correct wrist motility, although a natural skin color could not be achieved. Neither the Integra nor the ASE was rejected. Total wound coverage was achieved at 4 months, and completely normal skin was observed at 6 months.

Conclusions: This technique of combining an artificial dermal matrix with an ASE could be an alternative method to cover the donor sites of radial forearm flaps.
(ASEs)\textsuperscript{8,9} for the treatment of donor sites of radial forearm flaps.

**Materials and Method**

**ENGINEERING ASE**

Skin biopsies were obtained from the flap or the tracheostomy. All patients received detailed oral and written information, and an informed consent approved by the local institutional review board was obtained. Subsequently, the biopsies were processed as previously described.\textsuperscript{8,9} Briefly, the samples were divided into small fragments and subjected to 4 washes with phosphate buffered saline 5 mL containing trypsin and ethylenediaminetetraacetic acid (Gibco, Paisley, Invitrogen, UK) at concentrations of 0.05% and 0.02%, respectively. The resulting cell pellet was used to obtain the primary keratinocyte culture. The remaining tissue fragments were subjected to a fifth wash with collagenase (Sigma, Madrid, Spain) and Dulbecco’s modified Eagle’s medium (DMEM) at a concentration of 2 mg/mL to obtain a cell pellet for use in the primary fibroblast culture.

The keratinocyte growth medium was labeled as QN [DMEM (Gibco, Invitrogen)] and Ham F-12 (Gibco, Invitrogen) at a ratio of 3:1 plus 10% fetal bovine serum (Gibco, Victoria, Invitrogen, Australia) plus insulin (5 \( \mu \)g/mL; Sigma) plus cholera toxin (8 ng/mL; Sigma) plus adenine (24 mg/mL; Sigma) plus triiodothyronine (1.3 ng/mL; Sigma) plus hydrocortisone (0.4 \( \mu \)g/mL; Sigma) and required the use of lethally irradiated 3T3 cells as a feeder layer. For the cultivation of fibroblasts, DMEM supple-

![FIGURE 1. Top, Donor site before raising the flap. Middle, Bilayer artificial dermal matrix sheets are placed. Bottom, Grafting of an autologous skin equivalent over the new dermis. The autologous skin equivalent was easy to handle and had a thickness close to that of an ultrathin skin graft.](image)
mented with 10% fetal bovine serum was used. After 3 days of culture, epidermal growth factor (10 ng/mL; Austral Biologicals, San Ramon, CA) was added to the keratinocyte culture. When cells reached 90% confluence, a first pass was made into T-25 flasks for keratinocytes and into T-12.5 flasks for fibroblasts. The time until confluence for the first-pass keratinocytes was 7 to 8 days. During that period, a second pass was needed for fibroblasts into the T-75 flasks. Fibrin glue obtained from each patient’s blood sample was used for scaffolding. To construct a 75-cm² ASE, plasma 12 mL, fibroblasts $66 \times 10^5$, 1% calcium chloride in saline serum 2.1 mL, tranexamic acid 210 μL (FIDES Ecofarma, Almacera, Spain), and saline serum were used. The mixture was then allowed to solidify at 37°C for 30 to 60 minutes with QN medium 4.7 mL. After that period, keratinocytes were seeded. Fibroblasts were from the second pass and keratinocytes were from the first pass (one third of the total amount of the T-flask cells). The remaining cells were frozen as previously described.8 From a primary culture, 3 ASEs of 75 cm² each were obtained. To avoid keratinocyte differentiation, the submerged method was used. ASE handling became easier by following the method described by Meana et al.10

CASES AND SURGICAL PROCEDURES

Two cases of oral epidermoid carcinoma were selected. A radial forearm flap was used for reconstruction in these cases. In the first case, the flap measured 9 × 7 cm; in the second case, a 10 × 7-cm free flap was used. During surgery, a skin biopsy was taken from the tracheostomy (first case) or from the free flap (second case). In addition, a blood sample of 27 mL was taken. In the first case, a 10 × 12.5-cm bilayer artificial dermal matrix (Integra, Prim, Barcelona, Spain) sheet was used; in the second case, 2 5-cm × 5-cm bilayer sheets were applied.

The required periods before ASE grafting were 4 weeks in the first case and 3 weeks in the second case. An ASE was then grafted over the new dermis without the need for anesthesia. Sutures were not needed; instead, homogeneous compression with gauzes and bandage was applied for 7 days (Fig 1).

FIGURE 2. First case. Top, One week after grafting of the autologous skin equivalent, there are areas where clusters of mature cells seem to be developing (arrows). Bottom, Complete healing was achieved after 6 months.
Results

After 1 week, all ASEs successfully took hold. During the second week, the wound area became progressively smaller. Macroscopically, there were spots exhibiting more epidermal differentiation surrounded by the ASE; the authors believe stratification and differentiation were taking place in those areas. Vascularization also increased. After 2 months, the first case required a second ASE because a central defect persisted without epidermal coverage. A new ASE was designed using frozen cells from the patient, and after 4 months complete healing was achieved. In the second case, complete wound healing was accomplished at 4 months without the need for a new ASE (Figs 2, 3).

Appropriate functional and esthetic results were achieved, as was a correct mobilization of the wrist. One disadvantage observed was a color alteration of the regenerated skin, but this was not considered relevant by the patients (Fig 4).

No pain, infections, or other complications were recorded. Patients also showed normal cutaneous sensitivity in the grafted area.

Discussion

The method of choice to close donor sites after obtaining a radial forearm flap is primary closure with sutures or by a local cubital rotational flap. The cubital flap limits the width of the donor area to 6 cm and may present disadvantages such as chronic lymphedema and denervation of the volar forearm.

In cases in which the width is greater than 6 cm, the classic method to cover the donor site involves the use of a split-thickness skin graft, which causes major issues, such as pain and sensibility alterations in the graft donor site, tendinous exposure, and poor functional and esthetic results. To overcome these problems, different techniques have been used, such as the combination of ultrathin skin grafts with a de-epidermized acellular dermis or an artificial dermal matrix. The objective is to not only provide coverage, but also restore full functioning and appearance by adding epidermis and dermis to aid in wound healing.

Under this premise, the authors decided to use bilayer artificial dermal matrix sheets to obtain a new dermis, a fundamental component to obtaining good function and appearance in wound healing, and to combine it with the authors’ experience in tissue engineering. By following this method, neither a second surgery nor a second donor site was needed.
The ASE management was similar to that observed with split-thickness skin grafts. Adaptation to the forearm defects was easy because of its scaffold of fibrin glue; hence, sutures were not needed. In that sense, the procedure could proceed without anesthesia. Another advantage was cells could be frozen and used to construct another ASE, if needed.

The functional and esthetic outcomes were comparable to those observed by other investigators, with good wrist motility and skin elasticity. Appearance was good except for the skin color, a problem related to the use of the artificial dermal matrix described previously by others. The skin sensibility was normal.

A disadvantage worth mentioning could be the eventual necessity of a second ASE, as in the first case. Although patients did not report any discomfort, this requirement could be attributable to the fact that stem cells of the ASE mature early or that the cell volume in the ASE was not adequate to achieve complete healing, a hypothesis that needs to be studied in future researches.

The use of an artificial dermal matrix with an ASE in the donor site of a radial forearm flap is a novel approach, with less morbidity compared with other techniques. Even if a second ASE is required at a later stage, the low morbidity, the lack of need of a second donor site and additional surgery, the optimal functional outcome, and the acceptable esthetic results make this technique promising. Nevertheless, further studies with a larger number of patients are needed to validate the present findings.

References

FIGURE 4. Top left and right, Functional assessment was satisfactory, with a correct mobility of the wrist. Bottom left, Optimal skin flexibility was achieved from the dermal regeneration compared with outcomes when using split-thickness skin grafts alone.