

# Experiment assessment of mass effects in the rat: implications for small animal PET imaging



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## Abstract

*In vivo* imaging using positron emission tomography (PET) is important in the development of new radiopharmaceuticals in rodent animal models for use as biochemical probes, diagnostic agents, or in drug development. We have shown mathematically that, if small animal imaging studies in rodents are to have the same “quality” as human PET studies, the same number of coincidence events must be detected from a typical rodent imaging “voxel” as from the human imaging voxel. To achieve this using the same specific activity preparation, we show that roughly the same total amount of radiopharmaceutical must be given to a rodent as to a human subject. At high specific activities, the mass associated with human doses, when administered to a rodent, may not decrease the uptake of radioactivity at non saturable sites or sites where an enzyme has a high capacity for a substrate. However, in the case of binding sites of low density such as receptors, the increased mass injected could saturate the receptor and lead to physiologic effects and non-linear kinetics. Because of the importance of the mass injected for small animal PET imaging, we experimentally compared high and low mass preparations using *ex vivo* biodistribution and phosphorimaging of three compounds: 2-fluoro-2-deoxyglucos (FDG), 6-fluoro-L-metatyrosin (FMT) and one receptor-directed compound, the serotonin 5HT<sub>1A</sub> receptor ligand, trans-4-fluoro-N-{2-[4-(2-methoxyphenyl) piperazino]ethyl}-N-(2-pyridyl) cyclohexane- carboxamide (FCWAY). Changes in the mass injected per rat did not affect the distribution of FDG, FMT, and FCWAY in the range of 0.6–1.9 nmol per rat. Changes in the target to nontarget ratio were observed for injected masses of FCWAY in the range of ~5–50 nmol per rat. If the specific activity of such compounds and/or the sensitivity of small animal scanners are not increased relative to human studies, small animal PET imaging will not correctly portray the “true” tracer distribution. These difficulties will only be exacerbated in animals smaller than the rat, e.g., mice.

*Keywords* Animal imaging; PET; FDG; FMT; FCWAY

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## 1. Introduction

The driving force for the development of small animal PET scanners is threefold: the availability of various disease models presently available in small animals, the advantage of paired statistics when using serial imaging compared to serial autopsy, and the need to replace, reduce, and refine animal use [1]. Most of the animal models of neurological disease [2], cardiovascular disease [3], and cancer [4] are in rodents. The choice of the animal species is determined by the general knowledge of the neuroanatomy and neurochemistry, the ability to investigate behavioral responses and the cost to purchase and house the species. Rats and mice meet many of these conditions.

PET radiopharmaceuticals fall into three categories: (1)

those that trace nonsaturable systems (e.g. [<sup>18</sup>F] fluorid in bone), (2) intermediate saturable systems (e.g., 2-[<sup>18</sup>F] fluoro-2-deoxyglucos (FDG) to measure changes in glucose transporter (GLUT) and hexokinase activity, and 6-[<sup>18</sup>F]fluoro-metatyrosin (FMT) to measure dopamine metabolism), and (3) easily saturable, low-density systems (e.g., [<sup>18</sup>F] labeled FCWAY, a 5-HT<sub>1A</sub> antagonist to measure changes in 5-HT<sub>1A</sub> receptor density). Another example of the latter case is [<sup>18</sup>F] labeled dVIP, a radioligand to measure VIP receptor density in tumors [5].

These three classes of radiopharmaceuticals present increasing imaging challenges as the density of the target site decreases. If we assume that studies are performed in humans with minimal saturation of the target sites at the attainable specific activity, we can mathematically deduce the conditions necessary to obtain comparable images in rodents and then experimentally test the effect of these conclusions in rats using the three classes of radiopharma-

Table 1  
Properties of the compounds used for dissection studies.

Radiopharmaceuticals	mCi injected	Specific activity Ci/mmol*	nmol injected
2- <sup>18</sup> F]FDG	4	~1,000	4
2- <sup>14</sup> C]FDG	0.050	~0.3	167
2- <sup>18</sup> F]FDG	0.050	~1000	0.02
2- <sup>14</sup> C]FDG	0.050	~0.3	167
6- <sup>18</sup> F]FMT with carbidopa	0.050	0.49	102
[ <sup>18</sup> F]FCWAY	0.050	869	0.089

\* TOI = time of injection.

ceuticals. We show with a simple mathematical argument that comparable images are obtained only when the amount of radioactivity administered to the rat is roughly the same as to the human. Thus, we compared biodistribution data in rats obtained with milliCurie, or human, level doses of injected tracer (by autoradiography) to microCurie amounts of injected tracer (by dissection and well counting) to identify potential violations of the tracer principal within these classes. Studies using PiPET were also performed to show that comparable images can be obtained in the rat brain [6].

## 2. Materials and methods

### 2.1. Radiopharmaceuticals

The [<sup>18</sup>F]fluoride was taken directly from the <sup>18</sup>O target after proton irradiation and dissolved in phosphate buffer. The 2-<sup>18</sup>F]FDG was prepared by the Hamacher method [7]. The 6-<sup>18</sup>F]FMT was prepared following the publication of Namavari et al. [8]. To block aromatic amino acid decarboxylase (AAAD), carbidopa (5 mg/kg) was administered s.c. 30 min before 6-<sup>18</sup>F]FMT. The trans-<sup>18</sup>F]FCWAY was prepared by the method of Lang et al. [9].

### 2.2. Rat biodistribution studies

Adult male Sprague-Dawley rats (200–250 g) were injected intravenously with either 2-<sup>18</sup>F]FDG or trans-<sup>18</sup>F]FCWAY and intra-peritoneally with 6-<sup>18</sup>F]FMT (pre-injected with carbidopa as described above) (Table 1). The rats were sacrificed at times between 30 and 45 min post-injection of the F-18 labeled radiopharmaceutical and the brain was immediately placed in 0.3 M sucrose on ice. The brain was dissected on ice; the blood and various brain tissues were collected and weighed. The radioactive content of the blood and various tissues was assayed by gamma counting. Reported ratios were obtained using the DUR values (differential uptake ratio is %injected dose/g of target tissue ×body weight (g)/100).

Table 2  
Properties of the compounds used for autoradiography.

Radiopharmaceuticals	mCi injected	Specific activity Ci/mmol*	nmol injected
2- <sup>18</sup> F]FDG	4	~1,000	4
6- <sup>18</sup> F]FMT with carbidopa	3	0.60	5,000
[ <sup>18</sup> F]FCWAY	1	1266	1.0

\* TOI = time of injection.

### 2.3. Rat ex vivo autoradiography and data analysis

Adult male Sprague-Dawley rats (200–250g) were injected intravenously with the amount of radiopharmaceutical listed in Table 2 and sacrificed after 30 min. The carbidopa was pre-injected as described previously. The brains were removed, immediately quick frozen in dry ice, and cut into 20 μm slices using a BIHI cytomicrotome (Bright Instrument Co. and Hacker Instrument Inc, B2589). The slices were placed on slides, air dried, and placed on a phosphorimaging plate with a pixel size of 25 μm<sup>2</sup> (Fuji BAS-SR2025). After exposure the plates were scanned using a Fuji Bio-imaging Analysis System 5000. These plates have a high cross section for beta particles and relatively low cross section for gamma rays.

The quantitative data expressed as differential uptake ratio/m<sup>2</sup> of target tissue (DUR/m<sup>2</sup>) were determined by correlating cpm to PSL (photo-stimulated luminescence units). For each study adjacent brain slices at 6 different regions throughout the brain were used to determine the linear relationship of PSL to cpm (cpm= slope×PSL). One slice was exposed to the phosphorimaging plate overnight and the adjacent slice was counted on a 1480 Wallac Wizard gamma counter. Using the PSL units associated with the phosphorimaging slice and the cpm from the adjacent slice, the slope of the line was determined for each study (the slope varied from 0.169–0.192 between studies). Using this relationship, regions of interest could be converted from PSL/m<sup>2</sup> to cpm/m<sup>2</sup>, which was used to calculate DUR/m<sup>2</sup> by the following equation:

$$\text{DUR/m}^2 = \frac{(\text{cpm/m}^2) \times (\text{body weight (g)})}{\text{cpm injected} \times 100}$$

The area of interest on the phosphor plate was not multiplied by the slice thickness of 20 μm to give a measure of radioactivity per volume because of the additional assumptions concerning the efficiency of collecting positron decay at different depths.

### 2.4. Imaging using PiPET

As an adjunct to the in vitro measurements, we performed imaging studies in a group of rats using the PiPET (6) small animal PET imaging system, PiPET is comprised

Table 3  
Properties of radiopharmaceuticals used for PiPET imaging.

Radiopharmaceuticals	Sacrifice time (min)	mCi injected	Specific activity Ci/mmol*	nmol injected
[ <sup>18</sup> F]fluoride	120	1	~1,000	1
2-[ <sup>18</sup> F]FDG	45	4	~1,000	4
6-[ <sup>18</sup> F]FMT with carbidopa	45	4.4	0.70	6,298
[ <sup>18</sup> F]FCWAY				
Brain only	30	2.0	3,283	0.6

\* TOI = time of injection.

of two opposed bismuth germanate (BGO) detector modules 20-cm apart and operated in time coincidence. Each detector consists of a 26×22 array of 2 mm×2 mm×10 mm long BGO crystals coupled with optical grease to the face of a Hamamatsu R3941 position-sensitive photomultiplier tube (PSPMT). Each crystal is polished on five sides and finely ground on its entrance end. All crystals are double-wrapped on their long sides with PTFE tape including the entrance end. The field-of-view of PiPET is 55 mm (transverse)×45 mm (axial).

Tomographic imaging is performed by attaching the object to be imaged to a vertical rotation stage placed midway between the detectors. During data acquisition, the object is rotated continuously around this vertical axis to acquire all possible lines-of-response. Tomographic images are then reconstructed from lines that fall within a user-selected range of angles relative to the detector normals. The absolute central point source sensitivity of PiPET is 130 cps/ $\mu$ Ci with an isotropic reconstructed spatial resolution of 2.1 mm (FWHM) using filtered back-projection and ramp filter. Images shown in this report were reconstructed with the 3D OSEM resolution recovery algorithm (20 iterations) rather than FBP. Spatial resolution in these images is estimated at 1.5 mm. This scanner and its use have been described previously [6].

### 2.5. Rat PiPET studies and data analysis

Adult male Sprague-Dawley rats (200–250g) were injected with the amount of the <sup>18</sup>F labeled radiopharmaceuti-

icals and sacrificed at the times shown in Table 3. The animal was then attached to the rotation device so that the upper half of the animal was in the field-of-view and imaging begun. In the 6-[<sup>18</sup>F]FMT studies the rats were injected with carbidopa (s.c. 5 mg/kg) 30 min prior to the 6-[<sup>18</sup>F]FMT.

Target-to-non-target ratios were obtained from the PiPET images in each of these cases by defining three separate regions-of-interest in the target region and three in the reference region of each image. The regions were well within the borders of the chosen tissue. Ratios were computed from the average of the target values divided by the average of the non-target values. Each ratio represents the ratio from a single animal.

## 3. Results

Since a reference tissue is not available for 2-[<sup>18</sup>F]FDG and gray to white ratios are difficult to obtain, ratios of brain tissue to cerebellum for co-injected [<sup>14</sup>C]FDG and 2-[<sup>18</sup>F]FDG in rats were obtained by autoradiography and biodistribution studies (Table 4). The ratios were found to be similar whether the images were obtained using autoradiography with 50  $\mu$ Ci [<sup>14</sup>C]FDG or 4 mCi of 2-[<sup>18</sup>F]FDG (Table 4). Comparison of the ratios of the biodistribution studies using co-injected 10  $\mu$ Ci [<sup>14</sup>C]FDG or 100  $\mu$ Ci of 2-[<sup>18</sup>F]FDG were also similar although the absolute ratios had a higher trend. Since the injected mass of FDG is small compared to the concentration found in plasma (5 mM) and given the capacity of the glucose transporters (GLUT) and the velocity of hexokinase phosphorylation, this is not unexpected (see Sec. 4). The single distribution point is justified based on previous studies showing that the radioactivity in the brain is FDG 6-phosphate at this time point.

The mass effect of FCWAY and FMT was established over the range of 50 to 200 nmol per rat for FCWAY and 57 to 5645 nmol per rat for FMT (Tables 5 and 6). These data show that, at 50 and 200 nmol/rat WAY100635, binding is decreased significantly in the tissues studied, whereas FMT binding is not decreased over the experimental range.

Quantitation of 2-[<sup>18</sup>F]FDG, 6-[<sup>18</sup>F]FMT, and trans-[<sup>18</sup>F]FCWAY was carried out using autoradiography (Fig. 1b–1d) and *ex vivo* counting of dissected rat brain and ratios

Table 4  
Ratios of rat regional brain uptake to cerebellum (CB) for [<sup>18</sup>F]FDG and [<sup>14</sup>C]FDG (co-injected) determined by biodistribution and autoradiography at 30 min.

	Dose ( $\mu$ Ci)	Cortex/CB	Caudate/CB	Thalamus/CB	Hippocampus/CB	Brain stem/CB
[ <sup>18</sup> F]FDG	100	<b>1.37 <math>\pm</math> 0.05*</b>	<b>1.46 <math>\pm</math> 0.09</b>	<b>1.26 <math>\pm</math> 0.04</b>	1.03 $\pm$ 0.03	0.98 $\pm$ 0.03
[ <sup>14</sup> C]FDG	10	<b>1.43 <math>\pm</math> 0.02*</b>	<b>1.54 <math>\pm</math> 0.08</b>	<b>1.28 <math>\pm</math> 0.02</b>	1.05 $\pm$ 0.04	0.99 $\pm$ 0.04
[ <sup>18</sup> F]FDG	4000	<b>1.62 <math>\pm</math> 0.07†</b>	<b>1.69 <math>\pm</math> 0.09</b>	<b>1.48 <math>\pm</math> 0.15</b>	1.14 $\pm$ 0.10	1.29 $\pm$ 0.20
[ <sup>14</sup> C]FDG	50	<b>1.61 <math>\pm</math> 0.07†</b>	<b>1.65 <math>\pm</math> 0.10</b>	<b>1.47 <math>\pm</math> 0.11</b>	1.19 $\pm$ 0.03	1.24 $\pm$ 0.21

Values in bold are significantly different from one ( $p < 0.05$ )

\* Represents the mean ratio  $\pm$  SD by biodistribution;  $n = 5$ .

† Represents the mean ratio  $\pm$  SD by autoradiography;  $n = 3$ .

Table 5  
Mass effect on trans-[<sup>18</sup>F]FCWAY distribution (DUR) in rat brain.

[ <sup>18</sup> F]FCWAY	Cortex	Hippocampus	Cerebellum	Caudate
NCA*	2.01 ± 0.360	3.05 ± 0.422	0.160 ± 0.033	0.272 ± 0.064
+50 nmol <sup>†</sup>	0.564 ± 0.031	1.48 ± 0.141	0.101 ± 0.006	0.137 ± 0.020
+200 nmol <sup>†</sup>	0.217 ± 0.036	0.267 ± 0.040	0.168 ± 0.011	0.246 ± 0.064

Values represent the mean DUR ± SD, n = 5. The raw data are taken from Lang et al. J Med Chem. 1999 (9) and transformed to DUR values to correct for differences in animal weights.

\* No-carrier-added.

<sup>†</sup> Mass of WAY 100635 coinjected.

obtained for these three radiopharmaceuticals (Table 7). Tissues containing very low concentrations of receptor were used as a reference region.

FDG gave similar ratios using the two methods of analysis (Table 7). When 5 mg/kg carbidopa was injected before 6-[<sup>18</sup>F]FMT, the uptake in the striatum of the rat was clearly visualized. The ratio of uptake in the caudate to cerebellum was 1.7 using autoradiography. The ratio obtained by injecting 1/60 of the dose used in the imaging experiment and analyzing tissue radioactivity by gamma counting was also 1.7 at 45 min after injection (Table 8). The cerebellum is used as a reference tissue to normalize the study across experiments. The ratio in the target site of interest, the caudate, is the same in both experiments showing that at these mass differences, no mass effects were observed.

For trans-[<sup>18</sup>F]FCWAY, the hippocampus to cerebellum ratios obtained using biodistribution was 20, respectively while the hippocampus to cerebellum ratio was 19 by autoradiography (Table 7). Although the primary comparison in this study was between the low dose biodistribution studies and the high dose autoradiography studies, PiPET imaging was also carried out. PiPET images of the distributions of [<sup>18</sup>F]fluoride, 2-[<sup>18</sup>F]FDG, 6-[<sup>18</sup>F]FMT, and trans-[<sup>18</sup>F]FCWAY in the rat head and brain are shown in Figs. 1a–1d. Since the binding of fluoride by bone is known to be a non-saturable process, comparison with *ex vivo* autoradiography was not carried out. The very high target-to-non-target ratios, evident in the bone scan of the head, are virtually identical to those observed in the CT scan of the same animal (Fig 1a). PiPET images of 2-[<sup>18</sup>F]FDG, 6-[<sup>18</sup>F]FMT, and trans-[<sup>18</sup>F]FCWAY in the brain, shown in Figs. 1b–1d, gave similar ratios as those found by autoradiography although these were obtained from the average of three determinations in a single animal (data not shown).

PiPET has been used in small animal studies and found to be quantitative and of sufficient resolution for imaging dendritic cell migration [10], liver metabolism [11], and adrenal tumors [12].

#### 4. Discussion

Many studies on allometry suggest that the radiopharmaceutical injection be scaled either by the body weight or by the body surface area [13]. Many simple biological processes carried out in different species can be understood on the basis of these relationships. Thus, if weights were used as the allometric factor, the radioactivity injected into a rat would be scaled down by  $\sim 0.25 \text{ kg}/70 \text{ kg} = 1/280$ . With this calculation, the dose injected into a rat to maintain the same percentage saturation would be  $\sim 0.4\%$  of the human dose. If the resolution of the PET scanner required for animal brains were no greater than that required for humans ( $\sim 5 \text{ mm}$ ), this amount would be appropriate. However, that is clearly not the case.

It can be shown that although body weight scaling is appropriate if drug concentrations are to be the same in both human and animal, PET imaging in various species of different size requires a different relationship (Appendix I). If we wish to visualize the rat brain with the same relative acuity as we visualize the human brain, the volume of the resolution voxel in the rat must be shrunk approximately in proportion to the animal's mass relative to the human mass (1/280). Moreover, if a rat and human PET study are to be "equivalent" from an imaging point of view, the same coincidence rate must be observed from a rat resolution voxel as from a human resolution voxel. It follows, therefore, that the increase in dose with increasing resolution and the

Table 6  
Mass effect on 6-[<sup>18</sup>F]FMT distribution (DUR) in rat brain.

[ <sup>18</sup> F]FMT	Caudate	Thalamus	Cerebellum	Cortex
57 nmol*	0.724 ± 0.071	0.470 ± 0.047	0.399 ± 0.068	0.410 ± 0.048
597 nmol*	0.667 ± 0.119	0.476 ± 0.079	0.388 ± 0.057	0.399 ± 0.083
5645 nmol*	0.765 ± 0.119	0.555 ± 0.057	0.458 ± 0.050	0.410 ± 0.057

Values represent the mean DUR ± SD, n = 4–5.

\* Mass associated with [<sup>18</sup>F]FMT injected.

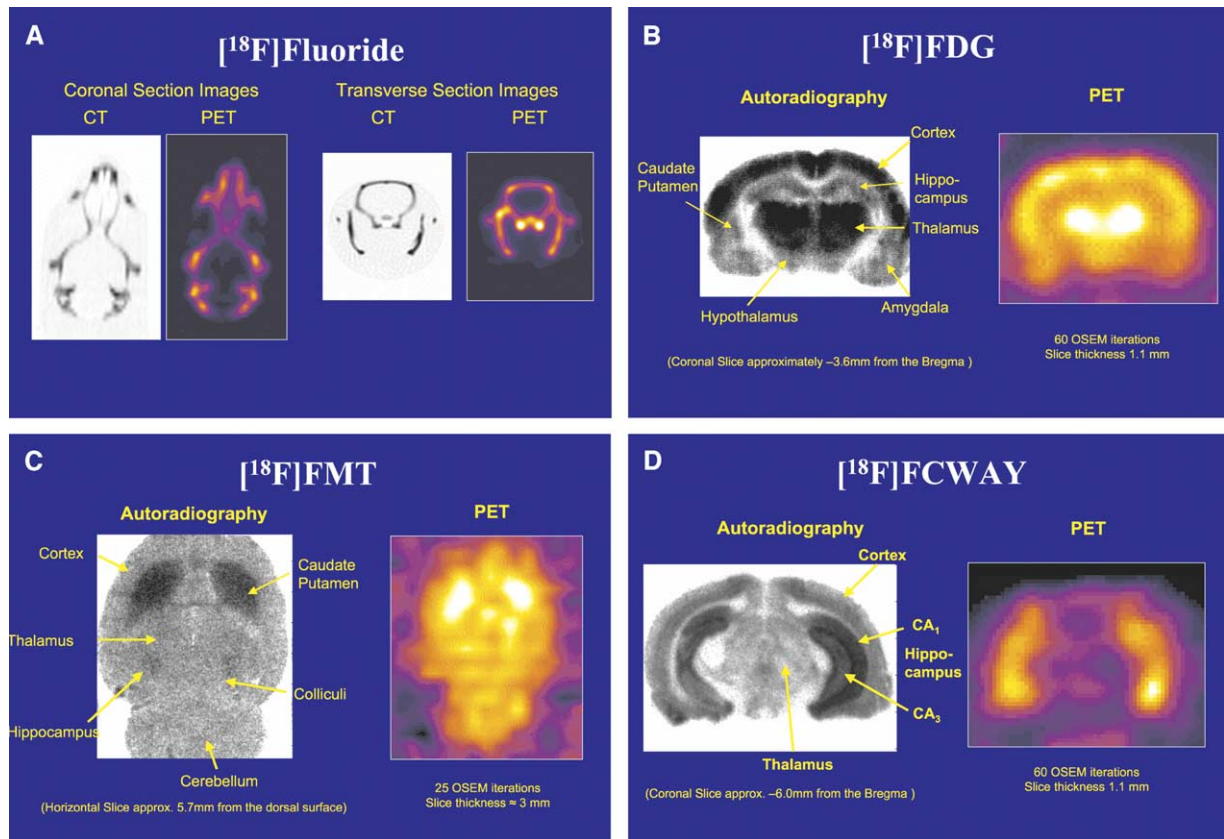


Fig. 1. Comparison of images of the rat head obtained by PET (right panels) in approximate spatial registration with images obtained by other methods in different animals (left panels): (A) CT versus F-18 fluoride-transverse section, (B) autoradiography versus F-18 FDG-transverse section, (C) autoradiography versus F-18 FMT-coronal section and (D) autoradiography versus F-18 FCWAY-transverse section. See Table 3 for details on the radiopharmaceutical dose.

decrease in dose with decreasing body weight cancel one another and the amount of tracer we must administer to the rat is about the same as to the human. It also follows that if a rat receives this human tracer dose, the concentration of drug in the animal will be higher in the ratio of human to rat body mass, potentially saturating vulnerable systems (the above argument ignores differences in attenuation, detection efficiency but which are included in the Appendix). Similar arguments can be made for brain imaging where the ratio in mass between humans and rat is approximately 1,000.

In humans, nonsaturable sites (e.g.  $^{18}\text{F}$  fluoride in bone) and intermediate saturable sites (e.g., 2- $^{18}\text{F}$ FDG to mea-

sure GLUT and hexokinase activity or 6- $^{18}\text{F}$ FMT) are not affected by low specific activities or large injected masses. No report of an effect of specific activity using  $^{18}\text{F}$  fluoride is reported and the United States Pharmacopoeia proposes the use of 2- $^{18}\text{F}$ fluoro-2-deoxyglucose at specific activities as low as 1 Ci/mmol for a 10 mCi dose of FDG. However, in easily saturated processes such as the binding of radiotracers to receptors, specific activity plays an important part.

In general, to avoid physiologic effects in the tracer study and to avoid decreasing the target to non-target ratio significantly, the receptor saturation should be below 5%. This can be seen from the equilibrium equation for binding attributed to Scatchard

Table 7  
Tissue ratios obtained by phosphorimaging compared to dissection data.

Radiopharmaceuticals	Tissue ratio used	Biodistribution	Autoradiography
$^{18}\text{F}$ FDG	Caudate to cerebellum	$1.5 \pm 0.1^*$	$1.7 \pm 0.1^\dagger$
6- $^{18}\text{F}$ FMT with carbidopa	Caudate to cerebellum	$1.8 \pm 0.03^*$	$1.7 \pm 0.1^\dagger$
$^{18}\text{F}$ FCWAY brain only	Hippocampus to cerebellum	<b><math>20.3 \pm 2.2</math></b>	<b><math>19.1 \pm 3.3^\ddagger</math></b>

Conditions given in Tables 1 and 2. Values in bold are significantly different from one ( $p < 0.05$ )

\* Mean ratio  $\pm$  SD;  $n = 5$ .

† Mean ratio  $\pm$  SD;  $n = 3$ .

‡ Mean ratio  $\pm$  SD;  $n = 4$ .

Table 8  
Ratio of tissue to cerebellum at 45 min after injection in rats at 0.050 mCi (biodistribution) and 3 mCi (autoradiography) of 6-<sup>18</sup>F]FMT.

	Dose ( $\mu$ Ci)	Cortex	Caudate	Brain stem	Thalamus	Hippocampus
<sup>18</sup> F]FMT	50	1.05 $\pm$ 0.05*	<b>1.69 <math>\pm</math> 0.08</b>	0.86 $\pm$ 0.43	1.16 $\pm$ 0.03	<b>1.20 <math>\pm</math> 0.05</b>
<sup>18</sup> F]FMT	3000	1.02 $\pm$ 0.04 <sup>†</sup>	<b>1.84 <math>\pm</math> 0.03</b>	1.10 $\pm$ 0.14	1.15 $\pm$ 0.01	1.13 $\pm$ 0.01

Values in bold are significantly different from one ( $p < 0.05$ ).

\* Represents the mean ratio  $\pm$  SD;  $n = 6$ .

<sup>†</sup> Represents the mean ratio  $\pm$  SD;  $n = 3$ .

$$B/F = (B_{\max} - B)/K_D \rightarrow \text{target to non-target ratio}$$

where the bound concentration over the free concentration ratio ( $B/F$ ) is related to the equilibrium dissociation constant ( $K_D$ ) and the total receptor concentration ( $B_{\max}$ ).

Although the kinetic analysis would be affected by increased mass, mostly by invalidating the assumption of first order kinetics used in most of the mathematical approaches, the single scan technique employed here will be affected as well. The  $B/F$ , which will be the bound concentration over the free concentration, is related to the maximum receptor concentration minus the bound concentration divided by the  $K_D$  and therefore if the bound concentration is large compared to the maximum receptor concentration, the observable  $B/F$  will be reduced. That equilibrium equation is the basis of why we have compared  $B/F$  at different specific activities and total mass injected. Although a mathematical approach to second order kinetics is clearly possible, the physiologic effects, as demonstrated by Bolo, et al. [15] for FDG, are something to be avoided in tracer experiments. Recent experiments with the muscarinic-2 receptor agonist, [<sup>18</sup>F]FP-TZTP, have also shown the physiologic effect of low specific activity studies [14].

If we now return to the examples of three classes of PET radiopharmaceuticals, we see that the uptake of [<sup>18</sup>F]fluoride in bone will not be affected if we inject 1000 times human dose per kilogram in a rat. However, the use of the human dose of 2-<sup>18</sup>F]FDG in rats may be more problematic if FDG is used at the USP limit of 1 Ci/mmol. Since the specific activity of FDG has been increased by  $>1,000$  with the adoption of the Hamacher synthesis for FDG [7], the chemical dose of FDG should not approach the saturation level of the enzyme involved. Bolo et al. [15] in studying the metabolic trapping efficiency of hexokinase in rabbit brain found that 70 and 210 mg/kg altered the pharmacokinetics in the brain compared to no-carrier-added [<sup>18</sup>F]FDG. Bolo et al. [15] saw saturation at 0.38 mmol (70 mg)/kg in rabbits. If saturation in rabbit brain is at  $>0.38$  mmol/kg and the maximal human dose is 140 nmol/kg, then there is a safety factor of  $\sim 2,700$  in the human brain. If  $1000\times$  the dose is injected in a rat, there will be a small safety factor using FDG prepared by electrophilic fluorination and a large safety factor for FDG prepared by the nucleophilic fluorination proposed by Hamacher et al. [7]. There has also been a series of studies using [<sup>19</sup>F]FDG *in vivo*. [<sup>19</sup>F]FDG is the NMR active, stable isotope of fluo-

rine. In order to obtain a measurable spectra with [<sup>19</sup>F]FDG and [<sup>19</sup>F]FDG 6-phosphate, the investigators used between 150 and 200 mg of [<sup>19</sup>F]FDG/kg. Physiologic effects were observed at  $>200$  mg/kg. Although the effect of mass paralleled that found in the [<sup>18</sup>F]FDG studies, the uptake of [<sup>18</sup>F]FDG and [<sup>19</sup>F]FDG was decreased by the added mass and the pharmacokinetics were altered [16].

A similar argument can be made for 6-<sup>18</sup>F]FMT, although it is prepared primarily using electrophilic fluorination at lower specific activities of 0.5–20 Ci/mmol. The capacity of the various enzymes in the metabolic pathway of FMT in the rat are high, e.g., the AAAD activity in rat plasma is 60 pmol/min/ml. Barrio et al. showed that even at 5 mg/kg carbidopa there was still peripheral AAAD metabolism [17]. They also showed that the rat striatum contained about half of the 6-<sup>18</sup>F]fluorotyramine found in the rat treated with carbidopa and very little 6-<sup>18</sup>F]FMT. The concentration of AAAD in the rat brain is very high, 106.7 nmol/min/g tissue [18]. In these situations where the radioligand is a substrate for a high capacity enzyme and the plasma concentration of amino acids is  $\sim 5$  mM, decreasing the uptake by saturating the target site is not likely. In fact, similar ratios of tissue to cerebellum (Table 8) was obtained with  $\mu$ Ci and mCi injections in rat for 6-<sup>18</sup>F]FMT. This is true for substrates for enzymes and transporters; inhibitors of enzymes and transporters would have to be treated like receptor binding radiotracers because the enzyme does not act on the radiotracer in a catalytic manner.

In the case of receptor binding radiotracers, the use of a  $1,000\times$  radioactive dose per kg for brain scanning could saturate the receptor above the 5% level. The calculations of saturation level for receptor binding radiotracers is very dependent on relatively small changes in specific activity, amount injected, and the percentage extracted by various brain regions [19]. For example, for trans-<sup>18</sup>F]FCWAY, the average 5-HT<sub>1A</sub> receptor concentration in the rat hippocampus is 80 nM [20]. With a 5 mCi injection of 1000 Ci/mmol specific activity at the time of injection, the mass injected would be 5 nmol. Based on the value of 1.3 %ID/g observed in the rat hippocampus, the concentration in a homogeneous solution would be 65 nM. Taking the nonspecific binding value of 0.06 %ID/g, the concentration of nonspecific binding would be 3 nM. Comparing the bound concentration (65 nM) with the hippocampal  $B_{\max}$  of 80 nM indicates that the hippocampal 5-HT<sub>1A</sub> receptor would be close to saturation.

A more rigorous calculation based on the equilibrium saturation binding with ligand depletion and assuming equal volumes of distribution indicates that the fraction of the total receptor occupied in the brain was 77% [21]. Motulsky has published a formula where he includes nonspecific binding [22]. This approach gave the same results in that the total receptor in the brain was near saturation. The 5-HT<sub>1A</sub> receptor concentrations in rats are similar to those found in humans [23].

In the actual PiPET study performed with FCWAY (Table 3), the average specific activity at time of injection was 3283 Ci/mmol and the amount injected was 2 mCi. The concentration of bound material using the same %ID/g in the hippocampus will be 0.6 nmol×0.013/g or 8 nM. This represents 10% of the total hippocampal receptor concentration whereas the previous calculation represented 77% for a mass injection of 5 nmol. Calculations using the approach put forward by Selikson [21] and Motulsky [22] also gave an occupancy of 10%. These calculations were based on a  $K_i$  for FCWAY of 1 nM. In various in vitro tests, we have obtained  $K_i$  values from 0.2 to 1 nM. Changing the  $K_i$  from 1 to 0.2 nM did not change the occupancy significantly. In the first example, a 5 mCi injection of 1000 Ci/mmol (5 nmol) resulted in a calculated receptor saturation of 77%. In the actual study, a 2 mCi dose of 3304 Ci/mmol; (0.6 nmol), the occupancy was reduced to 10%. It is clear from these examples that relatively small changes in dose resulted in major changes in occupancy.

There are two cautionary notes. The ratio of hippocampus to cerebellum in the biodistribution studies were comparable with that obtained with autoradiography (Table 7). This may be the result of the ability to analyze a very distinct region of the hippocampus with autoradiography versus the entire structure used for the biodistribution study. We have compared the PSL units in the CA1 band with the average PSL in the entire hippocampus from the autoradiography studies and found a ratio of ~5 indicating that regional differences were large.

Furthermore, the equations used here and those used by Hume et al. [19] and Meikle et al. [24] are equilibrium equations and there is no indication that FCWAY is at steady state in all tissues at 30 min after injection although the net efflux is similar in target tissue and metabolite corrected plasma [9]. In conclusion, small differences can change the percentage saturation substantially and experiments must be performed for each radiopharmaceutical at the injected dose and specific activity.

Hume et al. have carried out theoretical calculations based on the properties of three compounds with varying affinities for the limited number of target sites [19]. Hume et al. [19] show that in the range of dose observed for WAY 100635, the occupancy would be very dependent on the exact ED<sub>50</sub> value. Meikle et al. [24] have made a similar argument, but concentrated on tomographic sensitivity using average values for brain uptake and specific activities. Because small changes in affinity and receptor concentra-

tion have large effects on the occupancy, it is difficult to project a rule of thumb for various compounds as discussed above.

## 5. Conclusion

In present day practice we are far from obtaining the theoretical specific activity for <sup>18</sup>F or <sup>11</sup>C radiopharmaceuticals. However, for nonsaturable sites or high capacity sites injecting a human dose into a rat should not affect the pharmacokinetics. For receptor binding radioligands, the present experimental specific activities and the associated dose could lead to partial saturation of the target site in rats and more certainly in mice. An increase in the effective specific activity of the compound, an increase in the sensitivity of the animal scanner relative to human PET scanners, or both, will be required if new radiopharmaceuticals are to be evaluated in animal models by external imaging without these limitations. Although probably not sufficient for mice, FCWAY PET scanning in rats is possible with presently attainable specific activities and the sensitivity of contemporary small animal PET scanners.

## Appendix

The relationship between human and rodent imaging can be established by requiring that the same coincidence rate be observed from a rodent imaging "voxel" as from the human imaging voxel, where "voxel" is defined as the cube of the (isotropic) spatial resolution of the imaging device. If the subscript "r" refers to rodent and the subscript "h" refers to human, then equal voxel activities will occur when:

$$S_r C_r V_r \rho_r \exp[-u T_r] E_r = S_h C_h V_h \rho_h \exp[-u T_h] E_h \quad (1)$$

where  $S$ =specific activity ( $\mu$ Ci/gm),  $C$ =drug (radiopharmaceutical) concentration (gm/gm),  $V$ =voxel volume (cc),  $\rho$ =tissue density (gm/cc),  $u$ =linear attenuation coefficient of tissue at 511 keV (1/cm),  $T_h$  and  $T_r$  (in centimeters) are the characteristic thicknesses of tissue across the body at the location of the voxel of interest, and  $E$  is the fraction of coincidence pairs detected by the animal and human PET scanners compared to the number emitted from the respective voxels.

Solving Eq. (1) for  $C_r$  gives

$$C_r = [S_h/S_r] [V_h/V_r] [E_h/E_r] \exp[-u(T_h - T_r)] C_h \quad (2)$$

where we have assumed that the density of tissue in human and rat is the same. It is of interest to evaluate this expression for several relevant cases: when the same drug preparation is used in the rat as in the human, i.e. when  $S_r=S_h$  and in the case when  $C_r=C_h$ .

In either of these cases, the ratio of voxel volumes varies approximately as the ratio of body masses so that the volume ratio can be replaced by the ratio of human to rat body masses:  $70,000 \text{ gm}/250 \text{ gm}=280$ . If the transverse dimension of the human head is taken to be 18 cm and the rat head 3 cm, and the linear attenuation coefficient to be 0.08/cm, then the exponential term in Eq. (2) takes on the value: 0.3. In addition, current generation small animal PET scanner possess detection efficiencies not terribly different from human PET scanners so the efficiency ratio can also be set equal to unity. In the first case, therefore, the expression for the rat drug concentration becomes:

$$C_r = (280)(0.3)C_h \quad (3)$$

$$C_r = (84)C_h$$

That is, if the same drug, i.e., same specific activity, is used to image both rat and human, the drug concentration in the target tissue of the rat will be 84 times higher than in the human subject to obtain the same statistical image quality in the same imaging time and same relative degree of resolution as in the human study.

In the second case, the condition necessary to make the concentrations equal in both rat and human is

$$(S_r E_r)/(S_h E_h) = 84 \quad (4)$$

This relation implies that in order to make the concentrations equal, the specific activity-detection efficiency product for the rat study must be some 84 times greater than for the human study. This elevated ratio can be achieved by increasing the specific activity of the compound used in the rat study by this factor, by increasing the sensitivity of the small animal PET scanner by this factor or by increasing both such that the product of specific activity-detection efficiency is 84 for the rat study compared to the human study. Given that the detection efficiency of small animal PET scanners cannot be improved realistically by more than a factor of about five relative to human PET scanners, it follows that even in the rat some increase (a factor of 17 in this case) must be made in specific activity if the rat and human studies are to be performed at identical drug concentrations.

If the condition expressed in Eq. (4) is not met and the specific activity-detection efficiency ratio is the same in rats and humans, the total amount of radioactivity administered to the rat for imaging equivalency is given by the equation:

$$A_r = 84A_h[V_r/V_h] = (84)(250/70,000)A_h \quad (5)$$

$$A_r = 0.3A_h$$

where  $A$  is the total injected activity. Therefore, human and rat equivalency can be obtained if the rat receives 30% of the human dose. This dose, evidently, is not terribly different from the human dose, a result contrary to intuition and at odds with conventional allometry, which would predict a 280-fold reduction in injected activity for the rat. The cal-

ulation here assumes that when a human is given a human-appropriate dose, the tracer does not saturate the human system. The calculation also assumes that the duration of the human imaging study must also be matched by the animal study. Since the total number of recorded coincidences determines the statistical precision of the measurement of voxel activity, not just the coincidence rate, these conditions can be relaxed depending on the temporal behavior of the tracer. For example, a tracer that goes to the brain and remains fixed there can be imaged for extended periods at least in principle. In such a case, the dose and tissue concentration could be lowered in the rat by giving less activity but imaging for a longer time. The equivalency condition we have used here requires that tissue concentration, effective spatial resolution and total imaging time for the rat study are identical to the human study and that the human study exhibits satisfactory statistical precision of measurement under these conditions.

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